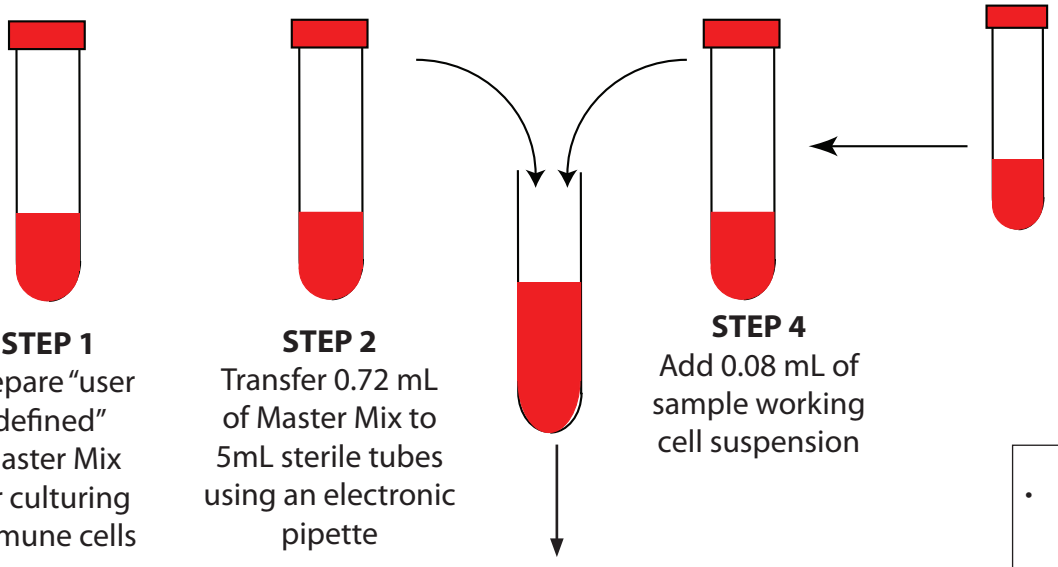
Quick Guide to ImmunoGlo™-96



STEP 1
Prepare "user defined" Master Mix for culturing immune cells

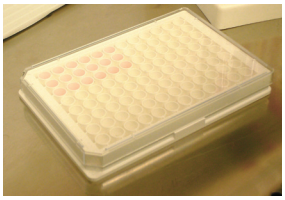
STEP 2
Transfer 0.72 mL of Master Mix to 5mL sterile tubes using an electronic pipette

STEP 3
Prepare cell suspension PBMNC or other immune cell fraction. Perform cell count and viability and adjust to working cell concentration

STEP 4
Add 0.08 mL of sample working cell suspension

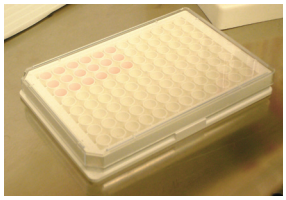
STEP 5
Gently mix by vortexing

STEP 6
Using an electronic pipette, dispense 0.1mL of Culture Master Mix into each of 6 replicate wells of the white culture plate



STEP 7
Incubate cells for specified time at 37°C in 5% CO_2 and 5% O_2 (preferred)

STEP 8
Calibrate and standardize proliferation assay prior to sample measurement (see separate Quick Guide)



STEP 9
Add 0.1mL of ATP-ER to each well, mix and measure bioluminescence after 10 min incubation in the dark

TIPS

- Always use at least a mononuclear cell (MNC) fraction as starting point.
- If possible, use self-calibrating electronic pipettes.
- Always dispense the Master Mix into the bottom of the well, never on the side.
- Always incubate plates in a humidified chamber to prevent drying out.
- Always use gloves when measuring ATP.
- Always perform calibration and standardization prior to measuring ATP samples.
- Always ensure well contents are mixed properly (see manual).
- Use sterile, adhesive foil (included) to maintain unused well sterility.

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