



A Step by Step Guide to using the HALO[®]-96 SPC-QC Kit: For Quality Control of Cord Blood, Peripheral Blood and Bone Marrow for Stem Cell Transplantation and Cord Blood Bank Processing Laboratories.











HALO[®]-96 SPC-QC is available using methyl cellulose or a liquid Suspension Expansion Culture (SEC)



We shall be using the SEC kit.









The single stem cell (CFC-GEMM) kit can be used for 16 cord blood, peripheral blood or bone marrow samples.



Everything you need for culture and ATP measurement is in the kit. Just add cells.

- We shall be using only 3 of the 16 Master Mix tubes for 3 cord blood samples.
- Remove the tubes and let the contents thaw.

- Prepare the cell suspension and adjust the concentration of each sample to 500,000 cells/ml.
- This is 100 times the final cell concentration/ well.

Remove 0.1ml of the cell suspension.

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Transfer the cell suspension from one sample to one of the HALO® Master Mix tubes containing 0.9ml of the culture reagents.

Mix the contents of each tube on a vortex mixer.

Now remove the sterile plate from the kit and take it out of the plastic foil.

Remove 0.1ml of the Culture Master Mix

Transfer 0.1ml of the Culture Master Mix to the first well of the 96-well plate.

The cell concentration is now further reduced 10 fold to 5,000 cells/ well.

Continue filling up 6 replicates of the row with 0.1ml of Culture Master Mix.

For the 3 samples we are testing, there will be 3 rows each with 6 x 0.1 ml replicates.

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Transfer the 96well plate to an incubator.

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Incubate the plate for 5 days at 37° C in a fully humidified atmosphere containing 5% CO₂ and, if possible, 5% O₂.

After 5 days incubation, we now prepare to measure ATP in the samples.

First, we have to perform the ATP standard curve.

An ATP standard and high and low controls are included with the kit.

Label 5 tubes: I, 0.5, 0.1, 0.05 and 0.01µM.

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Each ATP concentration is prepared by serial dilution: from I to 0.1 and then to $0.01 \mu M$; from 0.5 to $0.05\mu M$ using the medium provided with the kit.

Mix the contents of each tube prior to making the next ATP dilution.

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 Using the nonsterile, 96-well plate provided, first transfer
Iml of medium to the first 4 wells in a column.

Then transfer 0.1ml of each ATP dose to the next 4 vertical wells.

Start with the lowest ATP dose after the wells containing the medium.

The medium is the background.

The background (medium alone) and all 5 ATP doses have now been transferred to the nonsterile, 96-well plate.

Now transfer 0. I ml of the low control to each of 4 replicate wells in the last column.

Finish by repeating the procedure for the high control.

The kit contains one or more brown plastic bottles with ATP Monitoring Reagent (ATP-MR).

This contains a lysis reagent, luciferin and luciferase.

- Pour the reagent into a reservoir.
- Fix 8 tips to the pipette.
- Set the pipette to pick up 0.1ml of ATP-MR.

Transfer the ATP-MR to the wells of the first column.

Notice that the color changes when the ATP-MR is added.

Mix the contents of the wells using the same tips.

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Repeat the procedure for each column of well using new tips for each column of replicates.

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Leave the plate to stand for 2 minutes.

Then transfer the plate to the luminometer.

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Remove the plate lid and close the luminometer.

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The luminometer software can be programmed to calculate and plot the ATP standard curve.

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- The ATP standard curve is a linear regression using a log-log scale.
- The goodness of fit (r²) of the regression should be very near 1.

Once the ATP standard curve has been performed, remove the sample plate from the incubator.

Using only 6 pipette tips on the multi-channel pipette, add 0.1ml of the ATP-MR to each of the 6 replicate sample wells in the first row.

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After each addition of ATP-MR, mix the contents of each well using the same tips.

Repeat the procedure for each sample, changing the tips each time.

Incubate the plate at room temperature for 10 minutes.

- After 10 minutes, transfer the plate to the luminometer.
- Remove the lid from the plate.
- Start the instrument to read the luminescence.

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The Luminometer Readout

You may be able to customize your luminometer software to give you the output you want. This is an example of an exported SoftMax[®] file (Molecular Devices). Here you see a cord blood CFC-GEMM cell dose response (2,500, 5,000 and 10,000 cells/well) and the individual RLU values. The software has calculated the mean RLU and the mean RLU minus background (medium alone). It has also calculated the individual RLU values minus the background, followed by the mean standard deviation and percent coefficient of variation. From the ATP standard curve, it has then automatically calculated the individual ATP values and the mean, standard deviation and percent coefficients of variation. Alternatively, your luminometer software may already be programmed to give you this information.

Sample	Wells	Concentration	RLU Values	Mean RLU	Mean RLU-Bkg	RLU-Bkg	Mean RLU-Bkg	Std.Dev.	CV%	ATP values	Mean ATP	St. Dev ATP	%CV	
201	A7	2500	18.17	18.044	18	18.1	18	1.1	6	0.058	0.057	0.004	6.2	
	B7		18.453			18.4				0.059				
	C7		17.913			17.9				0.057				
	D7		16.566			16.5				0.053				
	E7		17.03			17				0.054				
	F7		17.571			17.5				0.056				
	G7		20.157			20.1				0.064				
	H7		18.491			18.5				0.059				
202	A8	5000	35.904	37.542	37.5	35.9	37.5	1.5	4	0.116	0.122	0.005	4.1	
	B8		37.029			37				0.12				
	C8		37.157			37.1				0.121				
	D8		40.406			40.4				0.131				
	E8		36.946			36.9				0.12				
	F8		37.519			37.5				0.122				
	G8		36.277			36.3				0.118				
	H8		39.1			39.1				0.127				
203	A9	10000	72.623	76.014	76	72.6	76	4.3	5.7	0.24	0.252	0.015	5.8	
	B9		83.959			83.9				0.279				
	C9		79.857			79.8				0.265				
	D9		70.192			70.2				0.232				
	E9		73.97			73.9				0.245				
	F9		76.643			76.6				0.254				
	G9		76.156			76.1				0.252				
	H9		74.713			74.7				0.247				

- These are the type of results you should expect.
 - Crisp, clean quantitative data with small coefficients of variation allowing excellent statistical analyses to be performed.

It saves time and money.

lt provides meaningful, quantitative results.

And if you do need help, we are here for you.

What HALO[®]-96 SPC-QC Format should you choose?

- You can choose between HALO[®]-96 MeC (methyl cellulose) or HALO[®]-96 SEC (Suspension Expansion Culture).
- In contrast to the former, HALO[®]-96 SEC SPC-QC:
 - Does not involve dispensing methyl cellulose.

- This produces lower errors and therefore lower CVs.
- It is faster and easier to perform.
- It is also 2 x more sensitive than HALO[®]-96 MeC SPC-QC.

Available for 4 applications

- Single stem cell (CFC-GEMM) kit: For routine QC.
- Duel stem cell kit: To detect both primitive (HPP-SP) and mature stem cells (CFC-GEMM).
- 4-Population Kit: To monitor the hematopoietic status of patients (CFC-GEMM, BFU-E, GM-CFC and Mk-CFC)
- 7-Population Kit: To monitor the lympho-hematopoietic status of patients (all of the above plus T-CFC, B-CFC and background).

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Order kits directly from HemoGenix[®] by: (1) Calling 719-264-6250 (2) Faxing 719-264-6253 (3) E-mailing info@hemogenix.com (4) Send a purchase order to: 1485 Garden of the Gods Road Suite 152 Colorado Springs, CO 80907

or

contact our distributors (see our website, <u>www.hemogenix.com</u> for details). Major credit cards accepted

